



CASE REPORT

Community-associated methicillin-resistant *Staphylococcus aureus* infection in Portugal[☆]

R. Nazareth^{a,*}, J. Gonçalves-Pereira^{a,d}, A. Tavares^c, M. Miragaia^c,
H. de Lencastre^{c,e}, J. Silvestre^{a,d}, P. Freitas^a, E. Gonçalves^b,
F. Martins^b, V. Mendes^a, C. Tapadinhas^{a,d}, P. Póvoa^{a,d}

^a Unidade Polivalente de Cuidados Intensivos, Hospital São Francisco Xavier, CHLO, Lisboa, Portugal

^b Laboratório de Microbiologia, Hospital São Francisco Xavier, CHLO, Lisboa, Portugal

^c Laboratório de Genética Molecular, Instituto de Tecnologia Química e Biológica (ITQB), Oeiras, Portugal

^d CEDOC, Faculdade de Ciências Médicas, Universidade Nova de Lisboa, Lisboa, Portugal

^e Laboratory of Microbiology, The Rockefeller University, New York, NY, USA

Received 14 February 2011; accepted 2 May 2011

KEYWORDS

Community-associated;
MRSA;
Staphylococcus aureus;
Necrotizing pneumonia;
Empyema

Abstract Methicillin-resistant *Staphylococcus aureus* (MRSA) has recently emerged as a cause of community-acquired infections among individuals without risk factors. Community-associated MRSA (CA-MRSA) appears to be more virulent, causing superficial mild skin and soft tissue infections to severe necrotizing fasciitis, and in rare cases, pneumonia.

Community-associated MRSA was first reported in Australia in the early 80s, after almost two decades in the USA, and then in several countries in Europe, Asia and South America. No data exists in Portugal.

We report the first case of CA-MRSA infection in Portugal, in a young adult with severe necrotizing pneumonia, complicated with bilateral empyema and respiratory failure.

© 2011 Sociedade Portuguesa de Pneumologia. Published by Elsevier España, S.L. All rights reserved.

PALAVRAS-CHAVE

comunidade associada;
MRSA;
Staphylococcus aureus;
pneumonia necrosante;
empiema

Pneumonia da comunidade por *Staphylococcus aureus* resistente à metilina em Portugal

Resumo Recentemente assistiu-se à emergência de infeções na comunidade por *Staphylococcus aureus* metilina-resistente (MRSA) em indivíduos sem fatores de risco. O MRSA associado à comunidade (CA-MRSA) parece ser mais virulento, causando desde infeções superficiais da pele e tecidos moles até fascite necrosante e, raramente, pneumonia.

O CA-MRSA foi inicialmente identificado na Austrália no início da década de 80 e, após cerca de duas décadas, surgiu nos EUA e em vários países da Europa, Ásia e América do Sul. Não existe informação disponível acerca da prevalência em Portugal.

[☆] Please cite this article as: Nazareth R, Infeção por *staphylococcus aureus* metilina-resistente da comunidade em Portugal. Rev Port Pneumol. 2012. Doi:10.1016/j.rppneu.2011.05.007

* Corresponding author.

E-mail address: raquelnazareth@gmail.com (R. Nazareth).

Os autores reportam o primeiro caso de infeção por CA-MRSA em Portugal, num adulto jovem com pneumonia necrotizante grave complicada por empiema bilateral e insuficiência respiratória.

© 2011 Sociedade Portuguesa de Pneumologia. Publicado por Elsevier España, S.L. Todos os direitos reservados.

Background

Staphylococcus aureus causes different types of infections, mostly minor skin and soft tissue infections (SSTI), but also severe pneumonia and sepsis, associated with high morbidity and mortality.¹

Methicillin-resistant *S. aureus* (MRSA), first reported in the United Kingdom in 1961,² is one of the most important agents of antibiotic-resistant health care-associated infections worldwide.³ In 2008, Portugal was the only European country with a prevalence of MRSA higher than 50%.⁴

During the past decade, community-associated MRSA (CA-MRSA) infections emerged worldwide among healthy individuals who did not have risk factors for hospital-acquired MRSA (HA-MRSA), mostly children and young adults.^{5,6} Both agents are epidemiological, genotypical and phenotypically unrelated.⁷ Community-associated MRSA was first reported in Australia in the early 80s,⁸ about a decade later in the USA⁵ and then in Europe, Asia and South America.⁹

Its exact prevalence is unknown. Outbreaks have been described in different settings in the community.¹⁰ A high prevalence was found in patients with SSTI,¹¹ namely intravenous drug users in North America,^{12,13} as well as in the UK.¹⁴ Initially, most of CA-MRSA isolates were only resistant to β -lactams, associated with the smallest types (IV and V) of the staphylococcal cassette chromosome *mec* (SCC*mec*).¹⁵ However, in recent years there has been increased resistance to other antibiotic classes.¹³ It frequently carries Panton–Valentine leukocidin (PVL) gene, an exotoxin causing leukocyte destruction and tissue necrosis, increasing severity of infections, especially pneumonia, with higher mortality rate.¹⁶

At present, the predominant CA-MRSA clones in the USA are USA300 (ST8-IVa) and USA400 (ST1-IV),¹⁷ particularly USA300, which is widely disseminated in the community and hospitals,¹⁸ whereas in Europe and Australia, the most relevant are the European clone (ST80-IVc)¹⁹ and the Southwestern Pacific clone (ST30-IV),²⁰ respectively.

Skin and soft tissue infections are the most common manifestations of CA-MRSA, but serious invasive infections, namely pneumonia, are also described.²¹ In a large USA study only 2% of CA-MRSA infections were pneumonia.²²

We describe the first documented case of CA-MRSA infection in Portugal, of a patient with severe necrotizing pneumonia complicated with bilateral empyema.

Case presentation

A 33-year-old homeless Armenian man, who had been living in Portugal for seven years, with a past medical history of chronic B and C hepatitis and active parenteral drug abuse came to the emergency department (ED) with a 5-day history of right pleuritic chest pain and, in the previous two weeks, non-quantified fever and purulent sputum.

The chest X-ray showed a lower bilateral pulmonary infiltrate and right-sided pleural effusion. HIV-check (4th generation ELISA method) was positive. A community-acquired pneumonia (CAP) with parapneumonic effusion was diagnosed and the patient was discharged on amoxicillin/clavulanate.

Two days later he returned to the ED with worsening symptoms. He was febrile, tachycardic and tachypneic, with pulse oxygen saturation of 95% on room air and had diminished breath sounds on the right hemithorax. Arterial blood gases revealed arterial partial pressure of oxygen (PaO₂) of 67.3 mmHg. Laboratory tests showed elevated C-reactive protein (35.4 mg/dL, which 2 days earlier had been 15.6 mg/dL) and leucocytosis ($14.5 \times 10^9/\mu\text{L}$; neutrophils 94%). No acid-fast bacilli were found on sputum smear (three samples). The patient was admitted to the Infectious Diseases ward. Clarithromycin was added and, since he had markedly depressed immunological state (46 CD4 T-cells/ μL , 10.3% with viral load $>180,000$ copies/mL), prophylactic therapy with trimethoprim–sulfamethoxazole was initiated. In the following 2 days, his clinical condition rapidly deteriorated with acute respiratory failure needing invasive mechanical ventilation. He was transferred to the intensive care unit (ICU).

In the ICU a bilateral thoracentesis was performed, revealing two large-volume empyemas. The chest CT scan showed right lung consolidation. Methicillin-resistant *S. aureus* was recovered from pleural fluid, bronchoalveolar lavage (BAL) and blood cultures. The microorganism was susceptible to vancomycin, with *in vitro* minimum inhibitory concentration (MIC) of 2 $\mu\text{g}/\text{mL}$ (Etest®) and resistant to ciprofloxacin, erythromycin and clindamycin; it was also susceptible to linezolid (MIC = 2 $\mu\text{g}/\text{mL}$), teicoplanin (MIC < 0.5 $\mu\text{g}/\text{mL}$), gentamicin (MIC < 0.5 $\mu\text{g}/\text{mL}$) and trimethoprim–sulfamethoxazole (MIC < 10 $\mu\text{g}/\text{mL}$). Vancomycin was started by continuous infusion adjusted to maintain serum levels of 20–25 mg/L. No improvement was noted after 7 days. A transesophageal echocardiogram excluded endocarditis.

Cultures were repeated revealing persistence of MRSA on BAL and tracheal secretions, with the same pattern of susceptibility. The hypothesis of CA-MRSA was suggested and linezolid IV was started. After a 10-day course there was complete clinical resolution.

The molecular characterization of the isolates was done by pulsed-field gel electrophoresis,²³ *spa* typing,²⁴ multilocus sequence typing²⁵ and SCCmec typing and subtyping.^{26,27} The presence of PVL was also assessed.²⁸ The etiological agent was identified as CA-MRSA belonging to the USA300 epidemic clone (ST8-IVa, t008, PVL positive).

The patient had a long ventilator course. A tracheostomy was performed on the 21st day. He was discharged from ICU on the 56th day and from hospital on the 179th day.

Conclusions

This is the first published case report of a CA-MRSA identified in Portugal, which led to a rare infection, necrotizing pneumonia with empyema.

The isolated CA-MRSA belongs to the epidemic clone USA300, which is wide spread in the USA, where it is also found in a hospital environment, being reported as the most common agent of SSTI.²⁹ It has also been isolated in Europe,⁹ but there were no prior cases reported in Portugal.

Most CA-MRSA infected patients reported have recent or concomitant influenza-like illness and/or recent antimicrobial exposure,³⁰ which was not the case with our patient. Nor did he have any other recognised risk factors for HA-MRSA, such as recent hospital admission, observation in a health care centre or recent antibiotic therapy. However, he was a homeless man with intravenous drug abuse habits, so in this context he could have had prior MRSA SSTI or exposure to another infected person. Not all cases have an identified risk factor like this one.

Necrotizing pneumonia associated with PVL producing strains has a high mortality rate that can reach 75%.¹⁶ In a series of 24 patients with CA-MRSA pneumonia, all patients with PVL producing strains died, compared with 47% of PVL non-producing strains (increased risk of 1.56).³¹ In another larger study, which included 50 patients with pneumonia caused by PVL producing CA-MRSA, the overall mortality was 56%.³² In our patient, CA-MRSA had elevated *in vitro* MIC to vancomycin and resistance to several antibiotic classes. Clinical deterioration may be explained by an increase in PVL levels, which can occur with sub-inhibitory concentrations of oxacillin and vancomycin.³³ In contrast, even sub-inhibitory concentrations of clindamycin and linezolid were shown to reduce PVL levels,³³ probably by inhibition of protein synthesis, which may explain the therapeutic success in our patient. Finally, others have shown that patients not responding to vancomycin frequently got better with the association of linezolid and rifampicin.³⁴ There are few available guidelines and no randomized clinical studies for comparing the different antibiotic therapies.

The recovery of a multidrug-resistant USA300 isolate from a drug user is worrying because of the high rate of transmission in this specific population,³⁵ causing concerns about possible increased rates of this agent in community-acquired infections in Portugal.

This report underlines the importance of CA-MRSA as a potential pathogen of severe CAP and the need to reinforce epidemiological surveillance. CA-MRSA infections do not cause specific signs or symptoms. However, it is crucial that attending clinicians and hospital microbiology laboratories have a high suspicion index to CA-MRSA when MRSA is isolated from patients with no risk factors for HA-MRSA but who have skin and soft tissue infections or pneumonia with a marked toxic and necrotizing component. Control efforts aimed at preventing the spread of the infection are feasible and can ultimately be successful.³⁶

Consent

Written informed consent was obtained from the patient for publication of this case report. A copy of the written consent is available for review by the Editor-in-Chief of *Portuguese Journal of Pulmonology*.

Author's contributions

RN drafted the manuscript. PF made substantial contribution by acquisition of data. JGP, JS, VM, CT and PP participated in the sequence alignment and also revised the manuscript critically for important intellectual content. EG and FM performed the microbiological assays and also revised the manuscript. AT, MM and MHL carried out the molecular genetic studies and also made substantial contribution in the manuscript's revision. JPG and PP have given final approval of the version to be published. All authors read and approved the final manuscript.

Conflicts of interest

The authors have no conflicts of interest to declare.

Acknowledgements

Molecular typing was supported by projects CONCORD (FP7-HEALTH-2007-B 222718) from the European Commission and project "Community-associated methicillin-resistant *Staphylococcus aureus* (CA-MRSA) in Portugal: a pilot study focusing on an emerging public health concern", from Fundação Calouste Gulbenkian, Portugal. A. Tavares was supported by grant SFRH/BD/44220/2008 from Fundação para a Ciência e a Tecnologia, Lisbon, Portugal.

References

1. Chambers HF, Deleo FR. Waves of resistance: *Staphylococcus aureus* in the antibiotic era. *Nat Rev Microbiol*. 2009;7:629–41.
2. Jevons MP. Celbenin'' – resistant *Staphylococci*. *BMJ*. 1961;1:124–5.
3. Kollef MH, Shorr A, Tabak YP, Gupta V, Liu LZ, Johannes RS. Epidemiology and outcomes of health-care-associated pneumonia: results from a large US database of culture-positive pneumonia. *Chest*. 2005;128:3854–62.
4. European antimicrobial surveillance system annual report; 2008. <http://www.rivm.nl/earss/database/>.
5. From the Centers for Disease Control, Prevention. Four pediatric deaths from community-acquired methicillin-resistant *Staphylococcus aureus*—Minnesota and North Dakota, 1997–1999. *JAMA*. 1999;282:1123–5.
6. Gorak EJ, Yamada SM, Brown JD. Community-acquired methicillin-resistant *Staphylococcus aureus* in hospitalized adults and children without known risk factors. *Clin Infect Dis*. 1999;29:797–800.
7. Mongkolrattanothai K, Boyle S, Kahana MD, Daum RS. Severe *Staphylococcus aureus* infections caused by clonally related community-acquired methicillin-susceptible and methicillin-resistant isolates. *Clin Infect Dis*. 2003;37:1050–8.
8. Saravolatz LD, Markowitz N, Arking L, Pohlod D, Fisher E. Methicillin-resistant *Staphylococcus aureus*. Epidemiologic observations during a community-acquired outbreak. *Ann Intern Med*. 1982;96:11–6.
9. Deleo FR, Otto M, Kreiswirth BN, Chambers HF. Community-associated methicillin-resistant *Staphylococcus aureus*. *Lancet*. 2010;375:1557–68.
10. Centers for Disease Control and Prevention (CDC). Outbreaks of community-associated methicillin-resistant *Staphylococcus aureus* skin infections—Los Angeles County, California, 2002–2003. *MMWR Morb Mortal Wkly Rep*. 2003;52:88.
11. Klevens RM, Morrison MA, Nadle J, Petit S, Gershman K, Ray S, et al. Invasive methicillin-resistant *Staphylococcus aureus* infections in the United States. *JAMA*. 2007;298:1763–71.
12. Huang H, Flynn NM, King JH, Monchaud C, Morita M, Cohen SH. Comparisons of community-associated methicillin-resistant *Staphylococcus aureus* (MRSA) and hospital-associated MRSA infections in Sacramento, California. *J Clin Microbiol*. 2006;44:2423–7.
13. Lloyd-Smith E, Hull MW, Tyndall MW, Zhang R, Wood E, Montaner JS, et al. Community-associated methicillin-resistant *Staphylococcus aureus* is prevalent in wounds of community-based injection drug users. *Epidemiol Infect*. 2010;138:713–20.
14. Otter JA, French GL. The emergence of community-associated methicillin-resistant *Staphylococcus aureus* at a London teaching hospital, 2000–2006. *Clin Microbiol Infect*. 2008;14:670–6.
15. Lee SM, Ender M, Adhikari R, Smith JM, Berger-Bachi B, Cook GM. Fitness cost of staphylococcal cassette chromosome *mec* in methicillin-resistant *Staphylococcus aureus* by way of continuous culture. *Antimicrob Agents Chemother*. 2007;51:1497–9.
16. Gillet Y, Issartel B, Vanhems P, Fournet JC, Lina G, Bes M, et al. Association between *Staphylococcus aureus* strains carrying gene for Panton–Valentine leukocidin and highly lethal necrotising pneumonia in young immunocompetent patients. *Lancet*. 2002;359:753–9.
17. McDougal LK, Steward CD, Killgore GE, Chaitram JM, McAllister SK, Tenover FC. Pulsed-field gel electrophoresis typing of oxacillin-resistant *Staphylococcus aureus* isolates from the United States: establishing a national database. *J Clin Microbiol*. 2003;41:5113–20.
18. Seybold U, Kourbatova EV, Johnson JG, Halvosa SJ, Wang YF, King MD, et al. Emergence of community-associated methicillin-resistant *Staphylococcus aureus* USA300 genotype as a major cause of health care-associated blood stream infections. *Clin Infect Dis*. 2006;42:647–56.
19. Larsen AR, Bocher S, Stegger M, Goering R, Pallesen LV, Skov R. Epidemiology of European community-associated methicillin-resistant *Staphylococcus aureus* clonal complex 80 type IV strains isolated in Denmark from 1993 to 2004. *J Clin Microbiol*. 2008;46:62–8.
20. Vlack S, Cox L, Peleg AY, Canuto C, Stewart C, Conlon A, et al. Carriage of methicillin-resistant *Staphylococcus aureus* in a Queensland Indigenous community. *Med J Aust*. 2006;184:556–9.
21. Francis JS, Doherty MC, Lopatin U, Johnston CP, Sinha G, Ross T, et al. Severe community-onset pneumonia in healthy adults caused by methicillin-resistant *Staphylococcus aureus* carrying the Panton–Valentine leukocidin genes. *Clin Infect Dis*. 2005;40:100–7.
22. Fridkin SK, Hageman JC, Morrison M, Sanza LT, Como-Sabetti K, Jernigan JA, et al. Methicillin-resistant *Staphylococcus aureus* disease in three communities. *N Engl J Med*. 2005;352:1436–44.
23. Chung M, de Lencastre H, Matthews P, Tomasz A, Adamsson I, Aires de Sousa M, et al. Molecular typing of methicillin-resistant *Staphylococcus aureus* by pulsed-field gel electrophoresis: comparison of results obtained in a multilaboratory effort using identical protocols and MRSA strains. *Microb Drug Resist*. 2000;6:189–98.
24. Aires-de-Sousa M, Boye K, de Lencastre H, Deplano A, Enright MC, Etienne J, et al. High interlaboratory reproducibility of DNA sequence-based typing of bacteria in a multicenter study. *J Clin Microbiol*. 2006;44:619–21.
25. Enright MC, Day NP, Davies CE, Peacock SJ, Spratt BG. Multilocus sequence typing for characterization of methicillin-resistant and methicillin-susceptible clones of *Staphylococcus aureus*. *J Clin Microbiol*. 2000;38:1008–15.
26. Milheirico C, Oliveira DC, de Lencastre H. Update to the multiplex PCR strategy for assignment of *mec* element types in *Staphylococcus aureus*. *Antimicrob Agents Chemother*. 2007;51:3374–7.
27. Milheirico C, Oliveira DC, de Lencastre H. Multiplex PCR strategy for subtyping the staphylococcal cassette chromosome *mec* type IV in methicillin-resistant *Staphylococcus aureus*: 'SCC*mec* IV multiplex'. *J Antimicrob Chemother*. 2007;60:42–8.
28. Lina G, Piemont Y, Godail-Gamot F, Bes M, Peter MO, Gauduchon V, et al. Involvement of Panton–Valentine leukocidin-producing *Staphylococcus aureus* in primary skin infections and pneumonia. *Clin Infect Dis*. 1999;29:1128–32.
29. Moran GJ, Krishnadasan A, Gorwitz RJ, Fosheim GE, McDougal LK, Carey RB, et al. Methicillin-resistant *S. aureus* infections among patients in the emergency department. *N Engl J Med*. 2006;355:666–74.
30. Wallin TR, Hern HG, Frazee BW. Community-associated methicillin-resistant *Staphylococcus aureus*. *Emerg Med Clin North Am*. 2008;26:431–55, ix.
31. Morgan MS. Diagnosis and treatment of Panton–Valentine leukocidin (PVL)-associated staphylococcal pneumonia. *Int J Antimicrob Agents*. 2007;30:289–96.

32. Gillet Y, Vanhems P, Lina G, Bes M, Vandenesch F, Floret D, et al. Factors predicting mortality in necrotizing community-acquired pneumonia caused by *Staphylococcus aureus* containing Panton–Valentine leukocidin. *Clin Infect Dis*. 2007;45:315–21.
33. Dumitrescu O, Boisset S, Badiou C, Bes M, Benito Y, Reverdy ME, et al. Effect of antibiotics on *Staphylococcus aureus* producing Panton–Valentine leukocidin. *Antimicrob Agents Chemother*. 2007;51:1515–9.
34. Micek ST, Dunne M, Kollef MH. Pleuropulmonary complications of Panton–Valentine leukocidin-positive community-acquired methicillin-resistant *Staphylococcus aureus*: importance of treatment with antimicrobials inhibiting exotoxin production. *Chest*. 2005;128:2732–8.
35. El-Sharif A, Ashour HM. Community-acquired methicillin-resistant *Staphylococcus aureus* (CA-MRSA) colonization and infection in intravenous and inhalational opiate drug abusers. *Exp Biol Med (Maywood)*. 2008;233:874–80.
36. Grundmann H, Aanensen DM, van den Wijngaard CC, Spratt BG, Harmsen D, Friedrich AW. Geographic distribution of *Staphylococcus aureus* causing invasive infections in Europe: a molecular-epidemiological analysis. *PLoS Med*. 2010;7:e1000215.