



LETTER TO THE EDITOR

Association between sputum myeloperoxidase concentration and acute exacerbation of bronchiectasis



Acute exacerbation (AE) signifies a crucial predictor for poor outcomes in bronchiectasis.¹ The major mechanism of AE in bronchiectasis is neutrophilic inflammation.² Consequently, modulating this pathway is crucial in controlling AE.²

Myeloperoxidase (MPO) is the most abundant protein in neutrophils, serving as a hallmark of neutrophil activation in respiratory bursts. Although neutrophil activation protects against microorganisms, exaggerated inflammation can damage alveolar cells.³ During extensive inflammation, neutrophils undergo secondary necrosis, releasing MPO that may damage resident lung cells. In chronic obstructive pulmonary disease, MPO plays a vital role in developing chronic inflammation,³ and its activity is linked to AE.⁴ However, the association between MPO concentrations and AE status in bronchiectasis remains unclear. Therefore, we aimed to determine whether sputum MPO concentrations reflect stable or AE status in bronchiectasis.

We prospectively enrolled 72 patients with bronchiectasis. All participants underwent sputum examination at the

time of study enrolment. Generally, patients in the stable group were followed up after 3 months, and those in the AE group were followed up after 1 to 3 weeks. The patient's condition was determined again at the second visit, and a sputum examination was performed. Patients were classified into four groups according to the absence or presence of AE (stable vs. AE) at the first and second visits (Fig. 1A). Among patients in the stable group at the first visit, patients who had a stable and AE status at the second visit were classified into the stable/stable and stable/AE groups, respectively. Similarly, among patients in the AE group at the first visit, patients who had a stable and AE status at the second visit were classified into the AE/stable and AE/AE groups, respectively. AE of bronchiectasis was defined as the worsening of ≥ 3 major symptoms lasting ≥ 48 hr, necessitating treatment change. The FACED and Bronchiectasis Severity Index (BSI) scores were used to assess bronchiectasis severity.⁵ The Sputum MPO function profile was determined by AnyLab F Myeloperoxidase (Z-Biotech, Korea), a point-of-care test (POCT) method based on immunofluorescence. Using a pipette, we added 3–5 μ L of phlegm obtained from the patient to the tube containing the detection buffer. After mixing the patient's sputum with the detection buffer, 100 μ L of the mixture was distributed into the sample well of the test cartridge

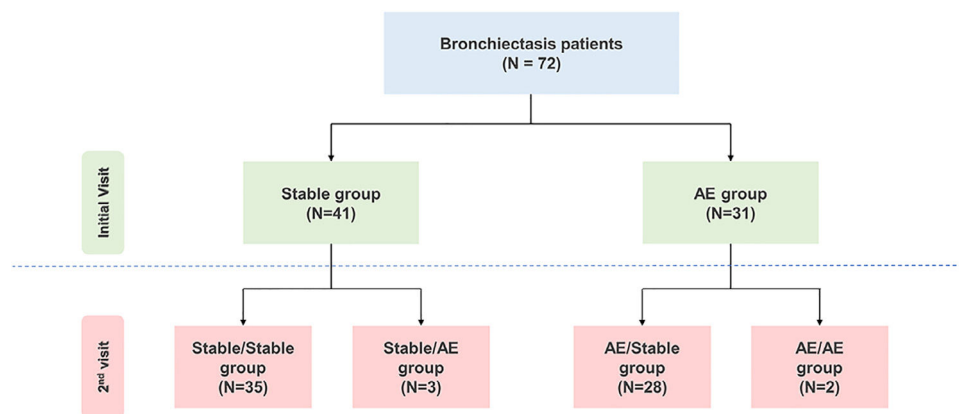


Fig. 1A Study population.

At the time of the initial visit, 3 patients in the stable group and 1 patient in the AE group did not visit the hospital at the second visit or the amount of sputum was small, and thus, sputum MPO concentrations could not be measured.

Abbreviations: AE, acute exacerbation.

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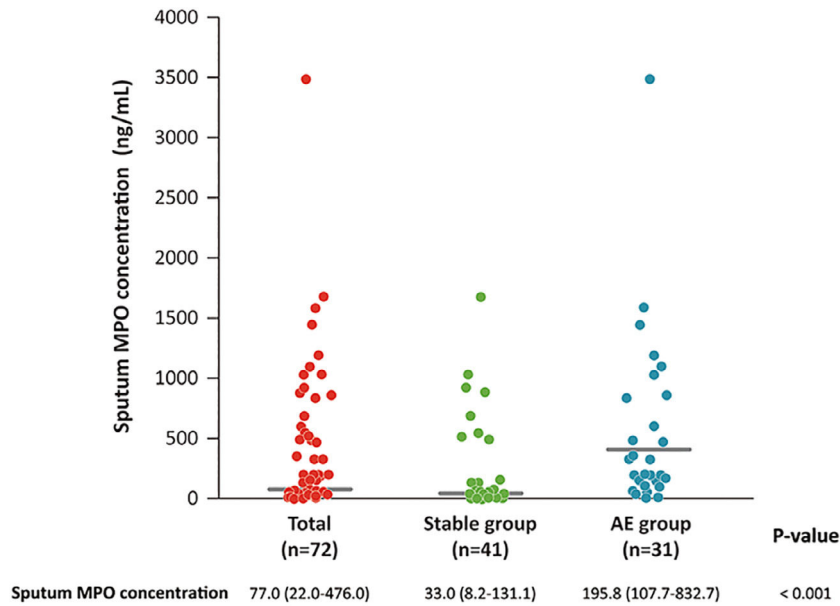


Fig. 1B Comparison of MPO concentrations between the stable and AE groups at the first visit. Abbreviations: AE, acute exacerbation; MPO, myeloperoxidase.

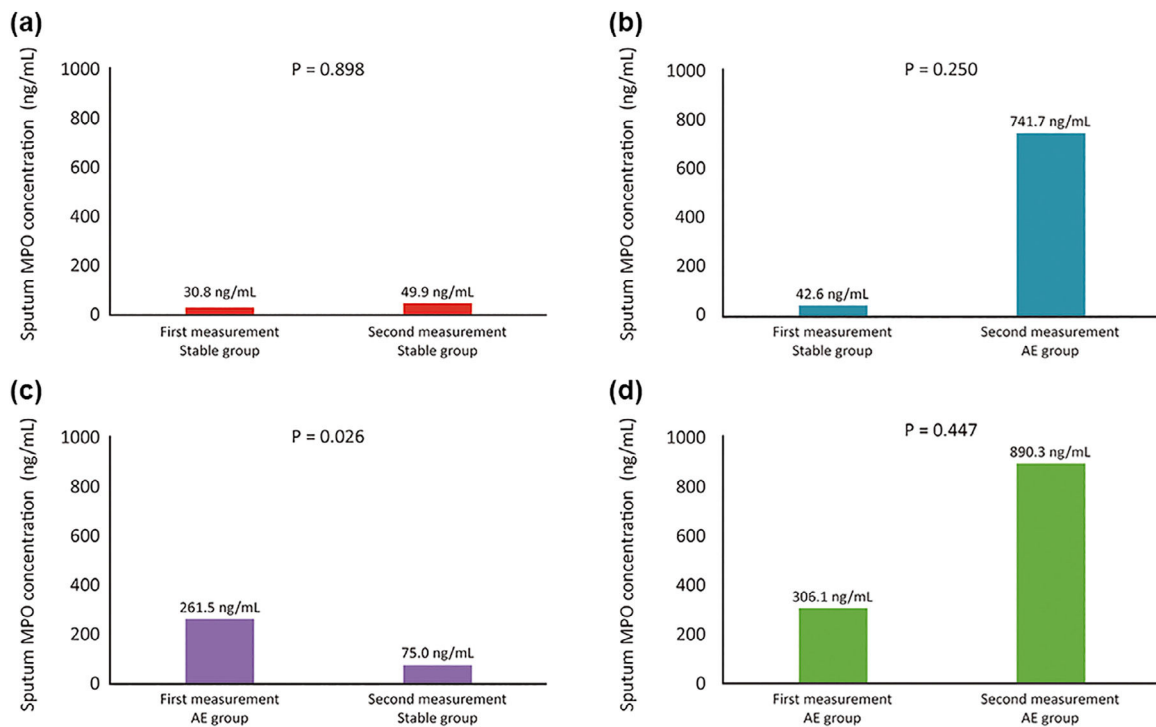


Fig. 1C Changes in MPO concentrations according to AE status of bronchiectasis.

- (a) Changes in MPO concentrations in the stable/stable group
- (b) Changes in MPO concentrations in the stable/AE group
- (c) Changes in MPO concentrations in the AE/stable group
- (d) Changes in MPO concentrations in the AE/AE group

Patients were classified into four groups according to the absence or presence of AE at the first and second visits as follows: stable/stable, stable/AE, AE/stable, and AE/AE groups.

Abbreviations: AE, acute exacerbation; MPO, myeloperoxidase.

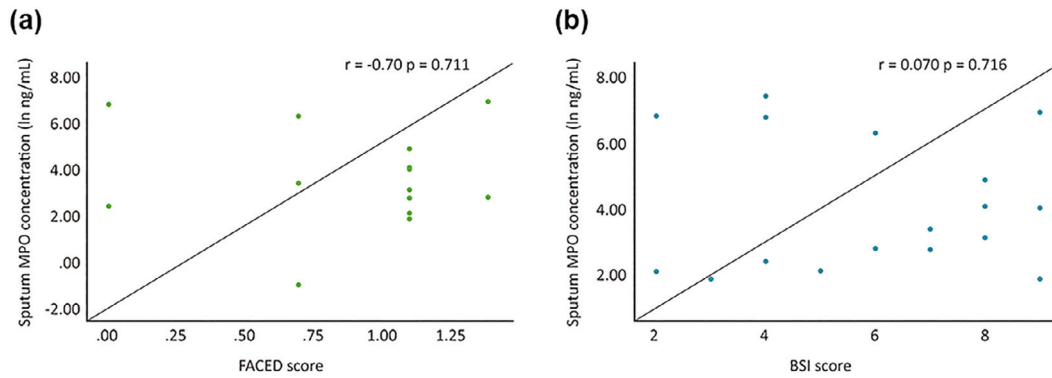


Fig. 1D Correlation between the disease severity of bronchiectasis and MPO concentrations in the stable group. Abbreviations: MPO, myeloperoxidase; BSI, bronchiectasis severity index; FACED, forced expiratory volume in 1 s (F), age (A), presence of chronic colonization by *Pseudomonas aeruginosa* (C), radiological extension (number of pulmonary lobes affected) (E), and dyspnoea (D).

and left at room temperature for 15 min. The results were obtained using the measuring device AnyLab F1.

The baseline characteristics of patients are shown in Table 1. Baseline MPO concentrations were higher in the AE group than in the stable group (median 195.8 ng/mL [interquartile range (IQR) 107.7–832.7] vs. 33.0 ng/mL [IQR 8.2–131.1], $p < 0.001$) (Fig. 1B). In the stable/stable group,

there was no significant change in MPO concentrations between the first and second measurements (median 30.8 ng/mL [IQR 6.4–218.0] vs. 49.9 ng/mL [IQR 15.8–231.7], $p = 0.898$). Conversely, in the stable/AE group, the second MPO concentration increased compared to the first MPO concentration (median 741.7 ng/mL [IQR 108.4–1194.3] vs. 42.6 ng/mL [IQR 22.4–59.0]) although it

Table 1 Baseline characteristics.

	Total (N = 72)	Stable group at the first visit (n = 41)	AE group at the first visit (n = 31)	p-value
Age, years	66 (59–72)	68 (60–75)	61 (58–69)	0.057
Sex, male	40 (55.6)	19 (48.7)	20 (64.5)	0.234
BMI, kg/m²	21.3 (19.5–23.3)	21.2 (18.8–22.5)	22.1 (20.1–23.7)	0.188
Smoking history				> 0.999
Never-smoker	50 (69.4)	28 (71.8)	22 (71.0)	
Current or ex-smoker	20 (27.8)	11 (26.8)	9 (29.0)	
Comorbidities				
Hypertension	10 (13.9)	6 (14.6)	4 (12.9)	> 0.999
COPD	24 (33.3)	14 (34.1)	10 (32.3)	> 0.999
Asthma	7 (9.7)	5 (12.2)	2 (6.5)	0.463
Diabetes mellitus	12 (16.7)	6 (14.6)	6 (19.4)	0.751
Cardiovascular disease	16 (22.2)	8 (19.5)	8 (25.8)	0.576
Neurologic disease	5 (6.9)	3 (7.3)	2 (6.5)	1.000
Malignancy	4 (5.6)	3 (7.3)	1 (3.2)	0.629
Connective tissue disease	6 (8.3)	4 (9.8)	2 (6.5)	0.693
History of tuberculosis	20 (27.8)	11 (26.8)	9 (29.0)	> 0.999
History of Pertussis	6 (8.3)	1 (2.4)	5 (16.1)	0.078
NTM-PD	18 (25.0)	12 (29.3)	6 (19.4)	0.416
Microbiology	35 (48.6)	18 (43.9)	17 (54.8)	0.476
<i>P. aeruginosa</i>	17 (23.6)	6 (14.6)	11 (35.5)	0.052
Others	18 (25.0)	12 (29.3)	6 (19.4)	0.416
Spirometry				
FVC, L	2.48 (2.07–2.98)	2.53 (1.99–3.13)	2.44 (2.17–2.96)	0.515
FVC, % predicted	72.5 (65.5–82.0)	72.0 (68.0–87.5)	73.0 (62.0–82.0)	0.724
FEV ₁ , L	1.78 (1.37–2.10)	1.86 (1.32–2.09)	1.76 (1.45–2.10)	0.935
FEV ₁ , % predicted	64.0 (53.3–75.5)	62.0 (52.0–71.5)	72.0 (55.0–80.0)	0.364
FEV ₁ /FVC	69.5 (59.0–72.8)	67.0 (55.0–72.5)	70.0 (63.0–74.0)	0.232

Table 1 (Continued)

	Total (N = 72)	Stable group at the first visit (n = 41)	AE group at the first visit (n = 31)	p-value
Modified Reiff score	8 (5–13)	8 (5–12)	8 (6–13)	0.347
mMRC	1 (1–2)	1 (1–2)	1 (0–2)	0.348
BSI score	8 (6–9)	8 (4–9)	7 (6–9)	0.994
FACED score	2 (1–3)	2 (1–3)	2 (1–3)	0.472
Lab findings				
WBC count, /mL	6950 (5940–8650)	7145 (5870–8463)	6570 (5945–9375)	0.809
Neutrophil count, /mL	4260 (3121–5980)	4270 (3185–4270)	4236 (3082–6034)	0.697
Albumin, g/dL	4.2 (4.0–4.5)	4.2 (4.0–4.4)	4.3 (4.0–4.5)	0.338
hs-CRP, mg/dL	0.4 (0.2–1.7)	0.3 (0.1–1.8)	0.4 (0.2–1.5)	0.340

Data are presented as medians (interquartile ranges) or numbers (%).

Abbreviations: AE, acute exacerbation; MPO, myeloperoxidase; BMI, body mass index; COPD, chronic obstructive pulmonary disease; NTM-PD, non-tuberculous mycobacterial pulmonary disease; FVC, forced vital capacity; FEV₁, forced expiratory volume in 1 s; mMRC, modified Medical Research Council; FACED, forced expiratory volume in 1 s (F), age (A), presence of chronic colonization by *Pseudomonas aeruginosa* (C), radiological extension (number of pulmonary lobes affected) (E), dyspnoea (D); BSI, Bronchiectasis Severity Index; WBC, white blood cell; hs-CRP, high-sensitivity C-reactive protein.

was not statistically significant (Fig. 1C). In the AE/stable group, the second MPO concentration decreased compared to the first MPO concentration (median 75.0 ng/mL [IQR 26.1–522.7] vs. 261.5 ng/mL [IQR 127.7–1042.9], $p=0.026$). Conversely, there was no significant change in the MPO concentrations in the AE/AE group (median 306.1 ng/mL [IQR 89.9–450.0] vs. 890.3 ng/mL [IQR 517.0–818.5], $p=0.447$) (Fig. 1C). As shown in Fig. 1D, there were no statistically significant correlations between MPO concentrations and the BSI and FACED scores in the stable group.

To the best of our knowledge, this is the first study to investigate whether changes in sputum MPO concentrations can reflect AE or stable status of bronchiectasis. In our study, the sputum MPO concentration was a biomarker related to AE status rather than disease severity. We observed no correlation between sputum MPO concentrations and BSI and FACED scores. Conversely, sputum MPO concentrations were significantly correlated with AE status.

Our study has several clinical implications. First, since we measured sputum MPO concentrations using the POCT method, our findings suggest that sputum MPO concentrations can be used as a POCT biomarker for evaluating the development of and recovery from AE in bronchiectasis. Utilising this method, clinicians can determine AE more objectively compared to the current symptoms-based diagnosis of AE. Second, MPO-guided assessment of AE of bronchiectasis may guide antibiotic prescriptions, potentially reducing unnecessary antibiotic use. Third, MPO could be a novel neutrophil-modulating therapeutic target for the treatment of AE in bronchiectasis.

This study has certain limitations. First, it was a single-centre study with a small sample size. Second, we quantitatively measured sputum MPO concentrations but did not evaluate sputum MPO activity. Thus, future studies are necessary regarding MPO activity and AE in bronchiectasis.

In conclusion, there was a significant correlation between changes in AE status and MPO concentrations. Our study suggests that MPO concentrations could be a useful biomarker that reflects AE in bronchiectasis.

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Declaration of Competing Interest

None.

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References

- Chalmers JD, Sethi S. Raising awareness of bronchiectasis in primary care: overview of diagnosis and management strategies in adults. *NPJ Prim Care Respir Med*. 2017;27(1):18.
- Chalmers JD, Haworth CS, Metersky ML, Loebinger MR, Blasi F, Sibila O, et al. Phase 2 trial of the DPP-1 inhibitor brensocatib in bronchiectasis. *N Engl J Med*. 2020;383(22):2127–37.
- Davies MJ, Hawkins CL. The role of myeloperoxidase in biomolecule modification, chronic inflammation, and disease. *Antioxid Redox Signal*. 2020;32(13):957–81.

4. Park HY, Man SF, Tashkin D, Wise RA, Connett JE, Anthonisen NA, et al. The relation of serum myeloperoxidase to disease progression and mortality in patients with chronic obstructive pulmonary disease (COPD). *PLoS One*. 2013;8(4):e61315.
5. Chalmers JD, Goeminne P, Aliberti S, McDonnell MJ, Lonni S, Davidson J, et al. The bronchiectasis severity index. An international derivation and validation study. *Am J Respir Crit Care Med*. 2014;189(5):576–85.

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